



Review

THE BONE-IMPLANT CONTACT AND OSSEOINTEGRATION OF DIFFERENT IMPLANT SURFACE TREATMENT: THE FINDINGS FROM A SYSTEMATIC REVIEW OF LITERATURE

F. Lorusso¹, G. Ascani², F. Inchingolo³, S. R. Tari¹, C. Bugea¹ and A. Scarano^{1,3}

¹Department of Innovative Technologies in Medicine & Dentistry, University of Chieti-Pescara, Italy;

²Department of Maxillofacial Surgery, Spirito Santo Hospital - Pescara, Italy;

³Department of Interdisciplinary Department of Medicine, "Aldo Moro" University of Bari, Bari, Italy;

⁴Visiting professor, Department of Oral Implantology, Dental Research Division, College Ingá, UNINGÁ, Cachoeiro de Itapemirim, Espirito Santo, Brazil

*Correspondence to:

Antonio Scarano, D.D.S., M.D.

Department of Innovative Technologies in Medicine & Dentistry,

University of Chieti-Pescara,

Via Dei Vestini 31,

66100 Chieti Italy

e-mail. ascarano@unich.it

ABSTRACT

The dental implant is associated with high long-term predictability for fixed rehabilitation in edentulous patients. The aim of the present review was to evaluate the state-of-art of dental implant surface treatment and their effect on osseointegration. The Pubmed/Medline, EMBASE, Cochrane Library databases has been screened to identify the histologic studies regarding the dental implant surfaces *in vivo*. The screening process revealed a total of 3173 papers with a total of 24 articles obtained by the manual search. A total of 482 duplicates have been removed and 2691 papers were assessed for the full-text evaluation. A total of 2527 articles were removed after the eligibility process and 149 articles were evaluated for the descriptive analysis. The implant osseointegration process is a complex combination of events that is oriented to an intimate interface between the dental implant surface and the host peri-implant tissues that oriented to produce a functional ankyrotic relationship between the components under the masticatory loading.

KEYWORDS: *implant, fixture, surface, osseointegration, bone*

INTRODUCTION

The dental implant osseointegration represents the turning point for edentulous ridge rehabilitations due to the more

Received: 02 August 2023

Accepted: 09 September 2023

ISSN: 2038-4106

Copyright © by BIOLIFE 2023

This publication and/or article is for individual use only and may not be further reproduced without written permission from the copyright holder. Unauthorized reproduction may result in financial and other penalties. **Disclosure: All authors report no conflicts of interest relevant to this article.**

recent advances in titanium biocompatibility, enhanced surface treatment and novel high hydrophilic/bioactive materials (1), with a long-term implant success rates over 90% (2). The osteoconduction process is involved with the recruitment and migration of osteogenic cells to the implant surface determines the early events correlated to the dental implant osseointegration. This phase produces a mineralised osteoid matrix deposition representing the main non-functionalised new bone formation at the level of the bone-implant interface. These events are strictly correlated with several factors including the dental implant microtopography (3). Other key factors are represented by the implant material, macro design, surface chemistry, bone density, surgical technique, and implant loading protocol (4). In literature, the bone-implant contact (BIC) percentage represent one of the most reliable parameter for dental implant osseointegration, while values >50% are considered optimal for a long term stability findings (5). On the contrary, the main disadvantage of this parameter is dynamic and could potentially vary over time. In addition, the BIC% is a bidimensional parameter that could be determined only with retrieved biopsies and is not replicable.

Also, the torque removal force has been suggested as an additional technique to assess the implant anchorage for research purposes evaluating the biomechanical behaviour of osseointegration (6). In this way, the roughness of a surface is one of the major factors contributing to implant stability, based on the assessment of the surface peaks and valleys. For this purpose, the arithmetic mean height deviation from a mean bi-dimensional plane (Ra); the Sa is considered in the case of a three-dimensional evaluation (7). The “osseointegration” concept was introduced by Branemark et al. (1) as the direct contact between living bone and a functionally loaded implant surface without interposed soft tissue at the light microscope level (8).

Today, titanium is the most common material for dental implants due to its low weight, high strength/weight ratio, low elasticity modulus, corrosion and wearing resistance, and biocompatibility (9). The most frequent titanium alloy (Ti6Al4V) is composed of 6% of aluminium and 4% vanadium (10). Lincks et al. (11) reported that the osteoblasts-like cells responded differently to cpTi and Ti6Al4V materials due to the alloy mosaicism and the surface chemistry. A passive surface oxide film around the titanium core (12) determines the interface generation between the titanium surface and the surrounding hard tissue. The oxide layer produces hydroxyl functional groups when exposed to the air environment (13). The hydroxyl functional groups dissociate when exposed to body fluid to generate an electric charge that is correlated to the pH of the fluids (13). In this way, the point of zero charge of rutile is 5.3, while the anatase point of zero charge is 6.2 (14, 15). The TiO₂ shows reported a neutral property. The hydroxyl concentration of TiO₂ is relatively large, representing an advantage for the proteins and cytokines adsorption promotion (12). The machined surfaces of the implant device are provided only by decontamination after the turning procedure.

Various treatments were proposed to improve the surface properties, taking advantage of rough interfaces with high implant stability and the surface contact area (6, 16, 17). In addition, rough surfaces seem to be effective in improving the osteogenic cell's behaviour (18, 19), proliferation and differentiation (20, 21) due to the release of signal mediators, transforming growth factor beta, and prostaglandin E2 (PGE2) (21-24). The optimal roughness for dental implant surfaces range is approximately 1.5 µm (25). Several methods have been suggested, such as modified surfaces, additive coating protocols, and subtractive methods, while today, the optimal surface type has not been defined. The present systematic reviews aimed to investigate the recent updates of bone-implant contact (BIC) effectiveness of different implant surface treatments.

MATERIALS AND METHODS

Article search methodology

The screening phase was conducted according to the Standards for Reporting Qualitative Research principles (SRQR) and the PRISMA guidelines (26). The selection was based on a keyword strategy synthesised in Table I.

Table I. Boolean search and keyword strategy.

Search Strategies	
Keywords	Advanced keywords search: ((dental AND (implant OR implants OR implantation OR implantology) AND (surface OR surfaces OR surface topography) AND (Histo*))
Databases	Pubmed/Medline, EMBASE, Cochrane Library

The papers' title and abstracts were assessed for an initial screening, and the manuscripts were limited to histological studies with bone-to-implant contact (BIC) outcomes. The full texts were finally collected and evaluated to assess the eligibility for the descriptive analysis.

Inclusion and exclusion criteria

The inclusion criteria for the eligibility synthesis were limited to histological studies that assessed bone-to-implant contact (BIC) outcomes from 1995 to today. The exclusion criteria were systematic and literature reviews, letters to the editor, *in vitro* and laboratory simulation, pilot studies, preliminary reports, no loading outcomes and early follow-up. The articles written in non-English language were excluded from the review.

RESULTS

Screening process

The electronic database identification process revealed a total of 3173 and 24 articles screened through a manual search. A total of 482 duplicates have been removed from the articles list, and 2691 articles have been submitted for the full-text screening process. A total of 2527 papers were excluded for the following reasons: 1302 for the wrong outcome, 668 for the wrong device, 259 for wrong study design, 147 for wrong publication type, 101 written in a foreign language, 34 for wrong study duration and 16 for the wrong study population.

Sandblasted surfaces

The sandblasting procedure was proposed by sandblasting the metal surface with gritting agents. The number and rotations speed, the flux pressure, and the granulometry of the agent particles (10, 27) determine the treatment. The sandblasting procedure increases the surface irregularity and the implant biomechanical characteristics. The most common sandblasting agents are aluminium oxide/alumina (Al₂O₃) and titanium oxide (TiO₂). The primary studies concerning the sandblasted surfaces are summarised in Table II. The procedure can influence the adhesion, proliferation, and differentiation of osteoblasts (20, 28).

Moreover, the fibroblasts result in a more difficult adhesion to the implant surface and a lower soft tissue proliferation around the implant in favour of the new bone formation (27, 29). Using surfaces blasted with Al₂O₃ particles was investigated compared to turned titanium surfaces. In the literature, the sandblasted implants showed higher BIC than the machined (30). In another study, the machined implants with Sa of 0.96 µm were compared to different blasting sizes, and after 12 weeks, all blasted surfaces demonstrated higher BIC compared with machined surfaces.

The blasting procedure leaves residual particles over the implant's surface, which can modify the bone healing process. Some authors support that the presence of remaining particles may benefit osseointegration, catalysing this process (31); others support that aluminium ions are suspected to impair bone formation by a possible competitive action to calcium (32-35). TiO₂ particle blasting was proposed to promote bone contact (27). Dental implants with TiO₂ surface were compared to machined implants with a statistically significant higher removal torque compared to machined implants. No differences in BIC were detected (25). A combination of TiO₂ blasted surface with fluoride ions has been proposed to improve the early osseointegration of dental implants (36). This method reported a bone-to-implant contact mean of >48% after 2 months of healing, which was higher than the blasting procedure alone (36).

At the same time, the precise nature of multinucleated giant cells is not thoroughly investigated, while a histological study suggested a priming effect on osteoblast activity similar to the hypothetical role of osteoclasts (37). Additional studies focus on sandblasted implants (38-40).

Plasma sprayed and plasma-chemical vapour surface

The plasma-sprayed treatments were studied in orthopaedics (41) and dental implants with no histological evidence of connective tissue infiltration at the interface level (42). Plasma-sprayed implants are obtained by spraying heat molten metal on the implant core, producing irregularly sized and shaped rounded particles and splats with valleys, pores and crevices (43). This treatment improves implant stability, bone growth (44), and higher surface contact area (10).

This treatment has been successfully investigated in rabbits (45), monkeys (46, 47), and humans (48-51), in different functional loading conditions (52). *In vivo*, no significant differences were detected between plasma-spray vs. machined implant, with a BIC percentage ranging between 55.9% and 56.2% (53). An alkali modification of the plasma-spraying

Table II. Comparative studies which used sandblasted implants.

Author	Implant surface	Results	Findings	Experimental design
Piattelli et al. (1998) ³⁰	(1) Al ₂ O ₃ blasted (2) Turned	BIC values (1) 60%±1.4% (2) 51%±1.9%	The blasted sites presented BIC values statistically higher in comparison to turned.	Implants inserted in the femoral articulation of rabbits. Healing period: 8 weeks
Piattelli et al. (1996) ³⁷	(1) Al ₂ O ₃ blasted (2) Turned (3) Plasma-spray	ACP, ALP activity (1), (2) and (3)	No MGS activity was reported for (1) and (2). At 2 weeks, Plasma spray revealed MGS activity.	Healing period: 2, 4 and weeks
Wennerberg et al. (1998) ³⁸	(1) Al ₂ O ₃ blasted (25µm, 75µm, and 250µm particles) (2) Turned	BIC values (1) Ranging from 31 to 47% (2) Ranging from 18 to 23%.	Blasted surfaces demonstrated more bone in contact to implant surface compared to turned surface.	Implants inserted in the tibia of rabbits. Healing period: 12 weeks
Wennerberg et al. (1996) ³¹	(1) Al ₂ O ₃ blasted (25 µm particles) (2) TiO ₂ blasted (25 µm particles)	BIC values (1) 49.2 % (2) 47.6 % Removal torque (1) 26.5 Ncm (2) 24.9 Ncm	No statistically different values concerning torque removal BIC values between the surfaces blasted with the same size of particles.	Implants inserted in the tibia of rabbits. Healing period: 12 weeks
Wennerberg et al. (1995) ²⁵	(1) TiO ₂ blasted (25 µm particles) (2) Turned	BIC values (1) 40.9 % (2) 34.5 % Removal torque (1) 35.4 Ncm (2) 29.2 Ncm	BIC values were not significantly different between the implants. However, TiO ₂ blasted implants demanded a statistically significant greater removal torque force than turned implants.	Implants inserted in the tibia of rabbits. Healing period: 12 weeks
Gottfredsen et al. (1992) ³⁹	(1) TiO ₂ blasted (10-53 µm particles) (2) Turned	Removal torque (1) 150 Ncm (2) 60 Ncm	BIC not significantly difference (data not shown), but, blasted implants presented higher removal torque values in comparison to turned sites.	Implants were immediately placed, in dogs. No prosthetic rehabilitation was performed. Healing period: 12 weeks
Ivanoff et al. (2001) ⁴⁰	(1) TiO ₂ blasted (25 µm particles) (2) Turned	BIC values (1) 37 % (2) 9 %	The analysis of the results revealed a significantly higher BIC for the blasted implants than turned groups.	Microimplants were inserted in the ridge of 27 patients. Mean healing period ranging from 3.9 to 6.3 months.
Rocci et al. (2008) ³⁶	(1) TiO ₂ blasted (25 µm particles) (2) TiO ₂ blasted with fluoride ions	BIC values (1) 24.8 % (2) 48.3 %	The implant surfaces grit-blasted seems to produce a positive effect on osseointegration, the adding of fluoride ions could produce a sensible bioactive effect on the integration process.	A total of 7 implants positioned in human mandible. Mean healing period 8 weeks.

technique by sodium hydroxide solutions at 40°C for 24 h can determine an oxide layer $R_a = 17.6 \mu\text{m}$) and 20 nm thickness (44). The main reported disadvantage of plasma-spray is the detachment of titanium after implant insertion. Franchi et al. (54) reported the particle detachment of plasma spray, sandblasted and acid-etched, and machined implants in sheep. The authors reported that the titanium particles were detected only in plasma-sprayed implants. This phenomenon can be related to the friction between the implant surface and host bone cavity during implant placement, but its implications are unclear. Recent non-thermal and argon-based plasma applications have been proposed for dental implants, reporting no significant changes in new bone formation compared to sandblasted dental implant (55-58).

On the contrary, a significant increase in argon-based plasma-spray implant-bone contact was reported by Qiao et al. compared to sandblasted and acid-etched fixtures (59). Several studies increased the new bone formation of hybrid titanium-zirconia dental implants obtained through a novel plasma spray technique (60, 61). The microwave plasma-chemical-vapour deposition (MWP-CVD) of diamond-coated Ti-Al6-V4 dental implants compared to Ti-Al6-V4 implants have been investigated (62, 63). No differences in BICs, delamination, or particle-dissociation due to shearing forces have been detected (62).

Acid-etched surfaces

The acid-etch implant was proposed to avoid the residues released from sandblasting, a non-uniform surface modification of the implant body (10). For this purpose, different acid-etching solutions have been proposed, such as chloridic (HCl), sulfuric (H₂SO₄), hydrofluoric (HF), and nitric (HNO₃), in different combinations. The acid-etching process effectiveness is by the baseline roughness, acid composition, temperature, and etching time. The histologic assessment results have been evaluated in Tables III and IV. A study compared two different etching of solution HCl and H₂SO₄, reporting that the surfaces presented a homogeneous distribution of small 1-2 μm peaks and valleys and a removal torque 4 times higher for acid etched (6). The dual acid-etched procedure was proposed to obtain a macro- and micro-texture of the titanium surfaces (6) and higher platelet and osteogenic molecular signals (64, 65). Degidi et al. (66) reported histologically a mean BIC percentage of 61.3%, with no gaps or fibrous tissues present at the interface. Similar BIC results were reported after four months of healing on non-loaded implants (67).

In immediate loading protocols, the mean BIC levels ranged between 78% and 85% in vivo in humans (68). In the posterior maxilla after 6 months of healing, the BIC values of dual acid-etched sites were statistically higher than in turned sites (~70%) (69). Different acid concentrations were evaluated by Cho et al. (70), reporting a removal torque for dual acid-etched implants statically higher compared to the machined surface. The removal torque of 2mm diameters triple-etched micro-implants has been investigated by Pontes et al. (71), who reported an increase of the strength resistance >6Ncm after 8 weeks of healing. In a sheep study, Jinno et al. (72) reported that the dual-acid etch technique produces similar BIC findings to dual etching-sandblasting surfaces. Some authors associated the main findings for bone response to the dental implant macro-geometry (72-74). Halldin et al. reported that nano- and microtopography indicted by dual etching can potentiate the initial biomechanical behaviour, while for a more extended osseointegration period, the surface interlocking capacity seems more effective (75). On the contrary, several studies reported that the roughness scale seems to be effective for new bone formation (76, 77).

A similar outcome was reported by Yoo et al. that highlighted higher BICs and removal torque resistance of dual-acid etched implants compared to grit blasted/acid etch with low bone remodelling rates (78). Also, others obtained similar results (79).

Sandblasted and acid-etched surfaces

The combination of sandblasting and acid-etching technique has been suggested to produce uniform scattered gaps and hole distribution and slightly less rough than the plasma-sprayed surface, which is characterised by profoundly irregular micro-texture and less favourable substrate for cell proliferation (80). The histological studies have been summarised in Tables V and VI. Higher torque removal values of sandblasted/acid-etch surfaces have been reported (+75%-125%) compared to acid-etched implants (81). Abrahamsson et al. (82) reported that the BIC values in dogs were significantly higher in sandblasted/acid-etched implants compared to machined surfaces. Similar results were observed in the comparative evaluation of sandblasted/acid-etched compared in plasma-sprayed implants(83). Sandblasted and acid-

Table III. Comparative studies which used acid etched and plasma-sprayed implants.

Author	Surface treatment	Results	Findings	Experimental design
Klokkevold et al. (1997) ⁶	(1) Acid-etched (HCl / H ₂ SO ₄)	Removal torque (1) 20.50 Ncm (2) 4.95 Ncm	The resistance to torque removal was 4 times greater for acid etched implants in comparison to the turned surfaces.	Implants were inserted in the femur of rabbits. Healing period: 2 months
	(2) Turned			
Cho et al. (2003) ⁷⁰	(1) Acid-etched (HF and HCl / H ₂ SO ₄)	Removal torque (1) 34.7 Ncm* (2) 15.2 Ncm	Dual acid etched implants required a higher removal torque average force than the turned surface implants.	Implants were inserted in the tibia of rabbits. Healing period: 12 weeks
	(2) Turned			
Weng et al. 2003 ⁶⁷	(1) Acid-etched (Osseotite [®])	BIC values (1) 62.5 % (2) 39.5 %	BIC values were significantly higher in dual acid-etched sites in comparison to turned sites.	Implants were inserted in areas with poor bone quality in the mandible of dogs. Healing period: 4 months
	(2) Turned (ICE [®])			
Klokkevold et al. (2001) ⁷⁹	(1) Acid-etched (HCl / H ₂ SO ₄)	Removal torque (1) 27.40 Ncm (2) 59.23 Ncm	Statistically significant differences were observed between acid-etched and turned implants, and between plasma-sprayed and turned implants. However, differences between acid etched and plasma-sprayed were not statistically different.	Implants were inserted in the femur of rabbits. Healing period: 3 months**
	(2) Plasma-spray	(3) 6.73 Ncm		
	(3) Turned			
Pontes et al (2015) ⁷¹	(1) Triple Acid-etched	Removal torque (1) 3.3 ± 1.7 Ncm (2) 2.2 ± 1.3 Ncm (3) 6.7 ± 1.4 Ncm	The triple acid etching can create a promising and efficient surface for the process of osseointegration.	Healing period: 8 weeks Implants were inserted in rats.
Rezende de Jesus et al. (2017) ⁷³	(1) Acid-etched	BIC values 2 weeks	Bone-to-implant contact and BD increased with time in both surface treatments implants	Implants were inserted in dogs Healing period: 2 and 4 weeks.
	(2) Sandblasted and Acid-etched	(1) 19.57±13.57% (2) 20.33±7.99% 4 weeks (1) 40.25 ± 9.45% (2) 42.80± 4.48%		
Carr et al. (2000) ⁵³	(1) Plasma-spray	BIC values (1) 55.9 %	No significant differences could be observed between groups concerning the BIC percentage.	Implants were inserted in the mandible of baboons. No prosthetic rehabilitation was performed. Healing period: 6 months.
	(2) Turned	(2) 56.2 %		

etched surfaces reported increased osteoconductive cell proliferation characteristics compared to plasma-spray implants (80, 84, 85). The histological findings of sandblasted/acid-etched reported after six months of healing in humans a mean BIC of 76.6 % (86). After 40 months, a 75.4 % BIC mean was observed on retrieved human implants (87).

Some studies reported sufficient bone volume and density that sandblasted/acid-etched surfaces can present a success rate of 99 % after two years (88). The combination of acid-etching and ZrO₂ particles sandblasting produces an increased bone deposition compared to plasma-sprayed and machined implants (54). Several authors reported that the depth and distribution of irregularities, the cavity morphology, and contaminating elements derived from the treatment procedures

Table IV. *Histologic studies in which acid etched implants were retrieved from humans.*

<i>Author</i>	<i>Surface treatment</i>	<i>Results</i>	<i>Findings</i>	<i>Experimental design</i>
Testori et al. (2001) ⁶⁸	Acid etched (Osseotite®)	BIC values ranging from 78% to 85%	Implants were successfully used in immediately loaded protocol.	Histologic analysis of two retrieved immediately loaded implants.
Degidi et al. (2003) ⁶⁶	Acid etched (HCl and H ₂ SO ₄)	Mean BIC value 61.3%	No gaps or fibrous tissues were observed at the interface.	Healing period: 4 months. Histologic analysis of two retrieved implants. No prosthetic rehabilitation was performed.
Trisi et al. (2002) ⁶⁹	(1) Acid etched (Osseotite®) (2) Turned	BIC values (1) 72.35 % (2) 35.32 %	BIC values in dual acid-etched sites were statistically higher than in turned sites.	Healing period: 6 months. Histologic analysis of implants inserted in the posterior maxilla of 11 patients. Healing period: 6 months.

play an important role in cell behaviour (89). In different animal study models, the sandblasted and acid-etched surfaces seem to produce in animals very similar BICs (~60%) compared to RBM, acid treatments and micro-arc procedures with no significant differences (90-96). At the same time, Marinho et al. reported a significantly higher new bone contact compared to the comparison of machined implant surfaces (97). Similar results were obtained by Buser et al. (98).

Nodized surfaces and micro-arc treatment

The oxidation technique has been proposed to modify the oxide layer properties and the surface biocompatibility (99), avoiding the deposit of grit particles (100). The anodised surfaces are obtained by a voltage application on the titanium surface in an electrolyte bath. The treated surface appeared with micro-pores of variable diameters without cytotoxicity (101). The removal torque of different thicknesses of anodised surfaces was investigated, which was significantly higher than that of smooth surfaces (99).

In the rabbit model, anodised, anodised and hydrothermally treated, and machined implants were investigated, reporting BIC values ranging between 40% and 50% and removal torque differences between the study groups (102). Authors reported that differentiation and calcification occurred on rough and smooth surfaces, indicating that the porous microstructure could enhance cell proliferation (43). In literature, it was demonstrated that the voltage for the anodising technique could produce a sensible influence on osseointegration properties, while the optimal value seems to be at ~550 V (103). In this way, the micro-roughness generated by anodic oxidation seems to significantly ameliorate BICs compared to sandblasted surfaces (104, 105) and machined implants (106). Moreover, using a super-hydrophilic surface of anodic oxidation implants has been proposed to potentiate this histological finding (107), while using biologically-derived triterpenoids adjuvant coating seems to produce no significant effect on this parameter (108).

In addition, the electrochemical anion sulphuric acid and phosphoric acid incorporation significantly affect BICs with an increase of ~200% histological bone contact (109). The micro-arc surface oxidation treatment has been proposed to improve the titanium dental implant. The biocompatibility of micro-arc oxidation has been tested by several authors, producing an acceleration and enhancement of the fixture's osseointegration (90, 110–113). Dundar et al. (90) reported similar BIC means (~60%) comparing different surfaces RBM, SLA, micro-arc, and sandblasted-micro-arc treatment with no significant difference.

Hydroxyapatite-coated surfaces and ceramic-coating implants

Hydroxyapatite implants have been studied to improve bone-implant fixation due to an increased osteoblast activity to this contact and adhesion, proliferation, and differentiation (114). Histological findings of hydroxyapatite implant

Table V. Comparative studies that used sandblasted and acid-etched implants.

Author	Surface treatment	Results	Findings	Experimental design
Abrahamsson et al. (2004) ⁸²	(1) Sand-blasted and acid-etched	-	BIC values (data not shown) were significantly greater in sandblasted and acid-etched sites than in turned surfaces	Implants were inserted in the mandible of dogs.
	(2) Turned			No prosthetic rehabilitation was performed. Healing period: 1, 2, 4, 6, 8 and 12 weeks.
Marinho et al. (2003) ⁹⁷	(1) Sand-blasted and acid-etched	-	The SLA surfaces revealed a higher bone response vs. machined surfaces.	Implants were inserted in rats. Healing period: 5, 15, 30, and 60 days
	(2) Turned			
Coelho et al (2011) ⁵⁷	(1) alumina-blasting	BIC values	No significant differences of BIC were detected at 4 weeks. An higher reoval torque was detected for RBM implants.	Implants were inserted in dogs. Healing period: 4, weeks.
	(2) biologic blasting	(1) 40.13± 2.54%		
	(3) plasma	(2) 37.23 ± 2.14%		
	(4) microblasted RBM	(3) 38.56 ± 2.49 %		
	(5) Sand-blasted and acid-etched (AB/AE)	(4) 39.65± 2.27% (5) 38.72± 1.44%		
Cochran et al. (1998) ⁸³	(1) Sand-blasted and acid-etched (250-500µm corundum particles, and etched with HCl / H ₂ SO ₄)	BIC values (1) 71.68 % (2) 58.88 %	The sandblasted and acid etched implants had a significantly greater BIC percentage than did the plasma-sprayed. However, no qualitative differences in bone tissue were observed between groups.	Implants were inserted in the mandible of dogs. Loading period: 12 months Healing period: 15 months
	(2) Plasma-sprayed			
Buser et al. (1999) ⁹⁸	(1) Sandblasted and acid-etched (0.25–0.50 µm particles, etched with HCl / H ₂ SO ₄)	Removal torque (1) 1.43 Ncm (2) 1.54 Ncm (3) 0.26 Ncm	Statistically significant differences were observed between sandblasted and acid-etched and turned implants, and between plasma-sprayed and turned implants. However, differences between sandblasted and acid etched and plasma-sprayed were not statistically different.	Implants were inserted in the maxilla of miniature pigs. No prosthetic rehabilitation was performed. Healing period: 12 weeks*
	(2) Plasma-sprayed			
	(3) Turned			

*Data from the 1st and 2nd healing periods were not included in this table.

Table VI. Histologic studies in which sandblasted and acid-etched implants were retrieved from humans.

Author	Surface treatment	Results	Findings	Experimental design
Hayakawa et al. (2002) ⁸⁶	Sandblasted and acid-etched (Straumann®)	BIC value 76.6 %	Bone surrounding the implant was uniformly and maturely structured.	Histologic analysis of one retrieved implant that was inserted in the palatal bone of the maxilla of a patient as anchorage for orthodontic treatment. Healing period: 6 months
Sakakura et al. (2005) ⁸⁷	Sandblasted and acid-etched	BIC value 75.4 %	The surrounding bone healed in a well-organized pattern and could not be differentiated from the original alveolus.	Histologic analysis of one retrieved implant of a patient. Loading period: 40 months

indicated a BIC range between 87.5%-97.4% (115). This coating technique reported high survival rates at medium- and long-term follow-ups (2, 116, 117). After 12 years of loading, the survival rate of hydroxyapatite implants was 93.2%, statistically increasing compared to titanium implants (2). After 10 years of loading, the hydroxyapatite implants reported a BIC range between 70.74%-86.23% (118).

Piattelli et al. (119) reported a localised chronic suppurative bone infection associated with peri-implantitis in a hydroxyapatite-coated implant, where the coating appeared detached from the titanium surface. Different methods can be used for hydroxyapatite coating, such as coating/sintering, electrophoretic deposition, immersion coating, hot isostatic pressing, solution deposition, sputter coating, and thermal spraying techniques (120). Hydroxyapatite plasma-spraying was indicated to combine the hydroxyapatite characteristics and the bone-implant mechanical interlock associated with the plasma-spraying procedure. Higher BIC values were reported for hydroxyapatite implants than titanium plasma-spray implants and machined fixtures (121). The Resorbable Blast Material (122) also known as the technique of ion-beam-assisted deposition (IBAD) (123, 124) has been proposed to improve the coating quality properties.

In vivo, the BIC values were significantly higher in IBAD surfaces compared to blasted and machined implants. The authors suggested that the advantages of the HA-coated implants in the early healing period could be apparent, while the separation or fracture of the coating layer could be prevented. However, the resorption needs to be further investigated (123). Svanborg et al. (125) investigated different hydroxyapatite (HA) nanocoating thicknesses on titanium grade Ti-6Al-4V implants of 15 mm in length and 3.85 mm in diameter in rabbits.

The single layer-HA coating reported a mean Sa 0.91 (0.20) μmm while the double layer-HA coating showed a mean Sa 0.77 (0.19) μmm (125). After 9 weeks of healing, the single layer-HA coating reported higher values of removal torque ($p < 0.05$) and at 2 weeks reported an increase of almost 5% of new bone formation compared with the control and the double layer-HA coating. After 9 weeks, the BIC for both groups was similar (~60%) (125). The advantage of ceramic-coating implants has been described due to the high osteoconductivity of the surfaces, while these techniques can produce a surface biofunctionalisation that can increase the implant osseointegration (126-167).

The surface functionalisation seems to maintain the implant roughness, while Jimbo et al. reported no significant differences between the smooth bioceramic surface and the rough bioceramic coated implants (142). In addition, other studies investigated different ceramic coatings such as calcium carbonate, ceramic brushite, glass fibres, phosphate-containing polymers, magnesium-containing polymers, and calcium-phosphate (126-167). Granato et al. investigated the coating thickness and demonstrated that the optimal Ca- and P-derived bioceramic coating layer ranged between 300-500

Table VII. Comparative studies which used anodized implants.

Author	Surface treatment	Results	Findings	Experimental design
Sul et al. (2002) ⁹⁹	(1) Anodized (oxide thickness approximately 200, 600, 800 or 1000 nm)	Removal torque (1) Ranging from 0.113 to 0.129 Nm	The preliminary results of this study suggest that the oxide thickness influence the bone tissue formation.	Implants were inserted in the tibia of rabbits. Healing period: 6 weeks
	(2) Turned (oxide thickness: 17.4 nm)	(2) 0.075 Nm		
Son et al. (2003) ^{*102}	(1) Anodized	Removal torque (1) 51.35 Ncm	Difference between groups was not statistically significant concerning removal torque and BIC values (data not shown).	Implants were inserted in the tibia of rabbits. Healing period: 12 weeks
	(2) Turned	(2) 35.28 Ncm		
Ivanoff et al (2003) ¹¹³	(1) Anodized	BIC values (1) 34 %	BIC values were statistically higher in oxidized than in turned sites.	Histologic analysis of implants inserted in the ridge of 20 patients. Mean healing period: 6.6 months
	(2) Turned	(2) 13 %		

* Data from the 1st healing period, and an experimental group were not included in this table.

nm (150). Moreover, the fluorapatite and heated-hydroxyapatite coatings present a decreased resorption rate compared to hydroxyapatite implant surfaces (167).

Thermal oxidation and heat surface treatment

The investigation of innovative procedures able to contrast surface wearing and successful bioactivity and osseointegration represents the current breakthrough in implantology. Thermal oxidation aims to create a highly crystalline oxide coating able to potentiate the interaction between the titanium surface and the host surrounding bone (168, 169). A 700°C exposure for 1 hour by a controlled furnace of Ti6Al4V alloy can induce the formation of a rutile oxide layer that could improve the osteoblast attachment on the implant surface in vitro (170). In addition, the heat treatment at 800°C in the air for 1 minute also seems to increase the BICs in vivo of acid significantly etch Ti6Al4V implants (171). The Al obtained similar results (2) (3) abrasive particle blasting with thermochemical treatment in minipigs compared to SLA (shot blasting surface) (172).

Quameya et al. reported that adding a supplemental fluoridic acid etch to the thermally oxidised surface did not significantly affect osseointegration compared to standard SLA surface implant (173). The heat-derived oxide layer has been studied by Kim et al. (174), which compared different oxide layer thicknesses of 20nm to 80nm and the additional treatment of CaP coating. The same authors detected no significant differences in BICs and ISQ at 5 weeks on dogs (174).

Zirconia implants and acrylic materials

Zirconia (zirconium oxide, ZrO₂) is a ceramic material purposed as dental implant material due to its biocompatibility, esthetic properties, and mechanical behaviour, which are better than alumina (60, 61, 175-188). Zirconia is reported to present a bone contact similar to titanium implants 189,190. The interface is composed of a proteoglycan layer that is thicker than titanium (191, 192). Zirconia implants are biocompatible, bioinert, and radiopaque, with high corrosion and wearing resistance, flexion and fracture (193-197).

In rabbits, the BIC value of zirconia implants was 68.4% after 4 weeks with no foreign bone reaction and fibrous tissue infiltration at the level of the interfaces (198). Loaded zirconia implants were evaluated in monkeys, with BIC values ranging between 66%-81% (199). Zirconia implants submitted to Al₂O₃ sandblasting were compared to titanium (Al₂O₃ sandblasting followed by H₂O₂ and HF etching reporting BIC values of 67.4% for zirconia, and 72.9% for titanium surfaces with no statistically significant differences (190). Various types of zirconia implants have been investigated in the literature, while the most investigated are yttria-stabilised tetragonal zirconia polycrystalline (3Y-TZP) and ceria-stabilised zirconia-alumina nanocomposite (NanoZr) (176).

Mijhatovic et al. investigated three different roughnesses of zirconia implants compared to sandblasted large grit and acid-etched titanium implants, showing no significant differences in total BICs after 10 weeks on dogs. The hybrid hydrophilic titanium-zirconium alloy (TiZr1317) revealed a lower removal torque at 2 weeks compared to standard titanium implants, while no differences were detected at 4 and 12 weeks. At 4 weeks, hybrid hydrophilic titanium-zirconium alloy (TiZr1317) showed significantly higher BICs in the marrow area of 19.25% (179). Very few studies investigated in vivo the properties of plastic and acrylic resin implants (200). Okamatsu et al. (200) studied the hybrid titanium-plastic implants and evaluated a homogeneous 150- to 250-nm acrylic layer coating. The authors reported new bone formation in the test and control groups, with no direct bone contact with the plastic implant.

UV and biologically functionalised surfaces

In literature, photodynamically functionalised implant surfaces have been investigated (201-204). Mehl et al. (201) reported no significant differences between BICs and ISQ in a split-mouth study model using a high-energy UV-irradiation in epicrestally titanium implants. On the contrary, a significant increase in removal torque and BICs was reported by Shen et al. (203). The authors evaluated a different combination of SLA-surfaces treated by UV-bactericidal irradiation at 15-20W, 0.1mW/Cm² and 0.2mW/Cm² (203). The UV photo-functionalisation seems effective, especially in the early phases of osseointegration (202), with a significant increase in bone contact and dental implant stability. This treatment seems to take a significant advantage when combined with biologically functionalised treatment with fibronectin and osteopontin (204, 205) due to a significant increase in the hydrophilicity of the surface.

DISCUSSION

Several authors investigated the biological properties of dental implant surfaces under *in vitro* conditions. At the same time, this kind of research is consistent in investigating the specific cell response, the clinical relevance of these results is discussible, and the development of long-term clinical evaluations is fundamental. Different implant topographies seem to influence the outcome of dental implants, but the magnitude and clinical relevance of this influence are still being investigated.

On the other hand, many studies are being published to investigate the viability of modified surfaces. Regarding the titanium alloy, a study performed in rabbits reported that the removal torque was statistically different after 6 months and 12 months, where the cpTi implants were significantly more stable. The BIC means presented no significant differences between the materials (206). In another investigation, cpTi and Ti6Al4V dental implants were positioned in baboons, reporting that BIC means were significantly higher in cpTi and Ti6Al4V implants, but differences after six months were not significantly different (207).

Even if the use of the alloy represents a mechanical advance compared to cpTi, biomechanical tests revealed that cpTi presented an increased stability. Moreover, the titanium implants, after the air exposure, can form an oxide layer all over the surface of 2–5 nm thickness. The oxide layer (208, 209) plays a key role in corrosion resistance, biocompatibility and implant osseointegration (210-212). The layer is mainly formed by TiO₂ (213), and the crystalline structure, the thickness and stability of this layer varies according to the surfaces of the implant (99, 214, 215).

Promising findings for dental implants concerning nitride titanium (TiN), nanostructured texture, laser-treated surfaces, and ceramic materials have been recently reported (188, 216). The nanostructured surfaces (1-100nm) could improve the early interface and bone-implant contact (217, 218). Authors reported that dogs presented a higher percentage of newly formed bone in contact with nanostructured implants than plasma-spray and machined implants (219, 220), and BIC values ranged between 55 and 96% in humans (221).

Nitride titanium (TiN) was proposed to produce a surface less susceptible to the ions release. For this purpose, the physical vapour deposition technique can produce a thin TiN layer (~1μ) for an osseointegration quality similar to standard titanium implants. This layer increases corrosion resistance, lower bacterial adhesion, and a golden aspect of the implant surface (222-227). The laser ablation is a reproducible procedure for a controlled, micron-sized surface with topographical features on the flanks of the threads. Lasered implants demonstrated significantly higher BIC and removal torque peaks than machined implants (228, 229). Calcium phosphate and ceramic coating are correlated to a high chemical bonding property, similar to hydroxyapatite (86). Biphasic calcium phosphate (162, 230, 231) or tricalcium phosphates have been investigated as implant coating (232, 233).

In conclusion, proper long-term studies have been published for TiO₂ surfaces, but other surfaces are documented with a medium-term follow-up period (234). While clinicians should consider that several new treatment surfaces are constantly purposed and currently available in the market, long-term findings are necessary to comprehend their long-term biological response.

REFERENCES

1. Brånemark PI, Breine U, Adell R, Hansson BO, Lindström J, Ohlsson Å. Intra-Osseous Anchorage of Dental Prostheses: I. *Experimental Studies. Scandinavian Journal of Plastic and Reconstructive Surgery*. 1969;3(2):81-100. doi:10.3109/02844316909036699
2. Schwartz-Arad D, Herzberg R, Levin L. Evaluation of Long-Term Implant Success. *Journal of Periodontology*. 2005;76(10):1623-1628. doi:10.1902/jop.2005.76.10.1623
3. Davies JE. Understanding peri-implant endosseous healing. *Journal of Dental Education*. 2003;67(8):932-949.
4. Albrektsson T, Brånemark PI, Hansson HA, Lindström J. Osseointegrated titanium implants: requirements for ensuring a long-lasting, direct bone-to-implant anchorage in man. *Acta Orthopaedica Scandinavica*. 1981;52(2):155-170.
5. Albrektsson T, Johansson C. Quantified bone tissue reactions to various metallic materials with reference to the so-called osseointegration concept. *The bone-biomaterial interface University of Toronto Press, Toronto*. Published online 1991:357-363.
6. Klokkevold PR, Nishimura RD, Adachi M, Caputo A. Osseointegration enhanced by chemical etching of the titanium surface. A

- torque removal study in the rabbit. *Clin Oral Implants Res.* 1997;8(6):442-447. doi:10.1034/j.1600-0501.1997.080601.x
7. Wennerberg A, Albrektsson T. Suggested guidelines for the topographic evaluation of implant surfaces. *International Journal of Oral & Maxillofacial Implants.* 2000;15(3).
 8. Jalbout Z, Tabourian G. *Glossary of Implant Dentistry.* International College of Oral Implantologists; 2004.
 9. Lautenschlager EP, Monaghan P. Titanium and titanium alloys as dental materials. *International dental journal.* 1993;43(3):245-253.
 10. Scarano A, Piattelli M. Superfici implantari. In: Novello G. *Implantologia pratica.* Coordenons: New Service International S.r.l. 1a ed. 2005, p. 21-32.
 11. Lincks J. Response of MG63 osteoblast-like cells to titanium and titanium alloy is dependent on surface roughness and composition. *Biomaterials.* 1998;19(23):2219-2232. doi:10.1016/S0142-9612(98)00144-6
 12. Hanawa T. Titanium-Tissue Interface Reaction and Its Control With Surface Treatment. *Front Bioeng Biotechnol.* 2019;7:170. doi:10.3389/fbioe.2019.00170
 13. Cai K, Frant M, Bossert J, Hildebrand G, Liefelth K, Jandt KD. Surface functionalised titanium thin films: zeta-potential, protein adsorption and cell proliferation. *Colloids Surf B Biointerfaces.* 2006;50(1):1-8. doi:10.1016/j.colsurfb.2006.03.016
 14. Scarano A, Piattelli A, Polimeni A, Di Iorio D, Carinci F. Bacterial adhesion on commercially pure titanium and anatase-coated titanium healing screws: an in vivo human study. *Journal of Periodontology.* 2010;81(10):1466-1471. doi:10.1902/jop.2010.100061
 15. Scarano A, Tripodi D, Carinci F, Piccolomini R, D'Ercole S. Biofilm formation on titanium alloy and anatase-Bactercline® coated titanium healing screws: An in vivo human study. *Journal of Osseointegration.* 2013;5(1):8-12.
 16. Kasemo B. Biocompatibility of titanium implants: Surface science aspects. *The Journal of Prosthetic Dentistry.* 1983;49(6):832-837. doi:10.1016/0022-3913(83)90359-1
 17. Thomas KA, Kay JF, Cook SD, Jarcho M. The effect of surface macrotexture and hydroxylapatite coating on the mechanical strengths and histologic profiles of titanium implant materials. *J Biomed Mater Res.* 1987;21(12):1395-1414. doi:10.1002/jbm.820211205
 18. Chehroudi B, Gould TRL, Brunette DM. Effects of a grooved titanium-coated implant surface on epithelial cell behavior in vitro and in vivo. *J Biomed Mater Res.* 1989;23(9):1067-1085. doi:10.1002/jbm.820230907
 19. Brunette DM. The effects of implant surface topography on the behavior of cells. *International Journal of Oral & Maxillofacial Implants.* 1988;3(4).
 20. Bowers KT, Keller JC, Randolph BA, Wick DG, Michaels CM. Optimisation of surface micromorphology for enhanced osteoblast responses in vitro. *Int J Oral Maxillofac Implants.* 1992;7(3):302-310.
 21. Boyan BD, Batzer R, Kieswetter K, et al. Titanium surface roughness alters responsiveness of MG63 osteoblast-like cells to 1 α ,25-(OH) $_2$ D $_3$. *J Biomed Mater Res.* 1998;39(1):77-85. doi:10.1002/(SICI)1097-4636(199801)39:1<77::AID-JBM10>3.0.CO;2-L
 22. Kieswetter K, Schwartz Z, Hummert TW, et al. Surface roughness modulates the local production of growth factors and cytokines by osteoblast-like MG-63 cells. *J Biomed Mater Res.* 1996;32(1):55-63. doi:10.1002/(SICI)1097-4636(199609)32:1<55::AID-JBM7>3.0.CO;2-O
 23. Doillon CJ, Silver FH, Berg RA. Fibroblast growth on a porous collagen sponge containing hyaluronic acid and fibronectin. *Biomaterials.* 1987;8(3):195-200. doi:10.1016/0142-9612(87)90063-9
 24. Martin JY, Schwartz Z, Hummert TW, et al. Effect of titanium surface roughness on proliferation, differentiation, and protein synthesis of human osteoblast-like cells (MG63). *J Biomed Mater Res.* 1995;29(3):389-401. doi:10.1002/jbm.820290314
 25. Wennerberg A, Albrektsson T, Andersson B, Krol JJ. A histomorphometric and removal torque study of screw-shaped titanium implants with three different surface topographies. *Clin Oral Implants Res.* 1995;6(1):24-30. doi:10.1034/j.1600-0501.1995.060103.x
 26. Hutton B, Salanti G, Caldwell DM, et al. The PRISMA extension statement for reporting of systematic reviews incorporating network meta-analyses of health care interventions: checklist and explanations. *Ann Intern Med.* 2015;162(11):777-784.

doi:10.7326/M14-2385

27. Wennerberg A. Experimental study of turned and grit-blasted screw-shaped implants with special emphasis on effects of blasting material and surface topography. *Biomaterials*. 1996;17(1):15-22. doi:10.1016/0142-9612(96)80750-2
28. Schwartz Z, Martin JY, Dean DD, Simpson J, Cochran DL, Boyan BD. Effect of titanium surface roughness on chondrocyte proliferation, matrix production, and differentiation depends on the state of cell maturation. *J Biomed Mater Res*. 1996;30(2):145-155. doi:10.1002/(SICI)1097-4636(199602)30:2<145::AID-JBM3>3.0.CO;2-R
29. Abron A, Hopfensperger M, Thompson J, Cooper LF. Evaluation of a predictive model for implant surface topography effects on early osseointegration in the rat tibia model. *The Journal of Prosthetic Dentistry*. 2001;85(1):40-46. doi:10.1067/mp.2001.112415
30. Piattelli A, Manzon L, Scarano A, Paolantonio M, Piattelli M. Histologic and histomorphometric analysis of the bone response to machined and sandblasted titanium implants: an experimental study in rabbits. *Int J Oral Maxillofac Implants*. 1998;13(6):805-810.
31. Wennerberg A, Albrektsson T, Andersson B. Bone tissue response to commercially pure titanium implants blasted with fine and coarse particles of aluminum oxide. *Int J Oral Maxillofac Implants*. 1996;11(1):38-45.
32. Blumenthal NC, Cosma V. Inhibition of apatite formation by titanium and vanadium ions. *Journal of biomedical materials research*. 1989;23(S13):13-22.
33. Savarino L, Cenni E, Stea S, et al. X-ray diffraction of newly formed bone close to alumina- or hydroxyapatite-coated femoral stem. *Biomaterials*. 1993;14(12):900-905. doi:10.1016/0142-9612(93)90131-K
34. Toni A, Lewis CG, Sudanese A, et al. Bone demineralisation induced by cementless alumina-coated femoral stems. *The Journal of Arthroplasty*. 1994;9(4):435-444. doi:10.1016/0883-5403(94)90055-8
35. Darvell BW, Samman N, Luk WK, Clark RKF, Tideman H. Contamination of titanium castings by aluminium oxide blasting. *Journal of Dentistry*. 1995;23(5):319-322. doi:10.1016/0300-5712(94)00003-X
36. Rocci M, Rocci A, Martignoni M, Albrektsson T, Barlattani A, Gargari M. Comparing the TiOblast and Osseospeed surfaces. Histomorphometric and histological analysis in humans. *Oral Implantol (Rome)*. 2008;1(1):34-42.
37. Piattelli A, Scarano A, Corigliano M, Piattelli M. Presence of multinucleated giant cells around machined, sandblasted and plasma-sprayed titanium implants: a histological and histochemical time-course study in rabbit. *Biomaterials*. 1996;17(21):2053-2058. doi:10.1016/0142-9612(96)00052-x
38. Wennerberg A, Hallgren C, Johansson C, Danelli S. A histomorphometric evaluation of screw-shaped implants each prepared with two surface roughnesses: A histomorphometric evaluation of screw-shaped implants. *Clinical Oral Implants Research*. 1998;9(1):11-19. doi:10.1034/j.1600-0501.1998.090102.x
39. Gotfredsen K, Nimb L, Hjørting-hansen E, Jensen JS, Holmén A. Histomorphometric and removal torque analysis for TiO₂ hyphen;blasted titanium implants. An experimental study on dogs.: Torque analysis for TiO₂ -blasted implants. *Clinical Oral Implants Research*. 1992;3(2):77-84. doi:10.1034/j.1600-0501.1992.030205.x
40. Ivanoff CJ, Widmark G, Hallgren C, Sennerby L, Wennerberg A. Histologic evaluation of the bone integration of TiO₂ blasted and turned titanium microimplants in humans: Bone integration of titanium implants. *Clinical Oral Implants Research*. 2001;12(2):128-134. doi:10.1034/j.1600-0501.2001.012002128.x
41. Hahn H, Palich W. Preliminary evaluation of porous metal surfaced titanium for orthopedic implants. *J Biomed Mater Res*. 1970;4(4):571-577. doi:10.1002/jbm.820040407
42. Schroeder A, van der Zypen E, Stich H, Sutter F. The reactions of bone, connective tissue, and epithelium to endosteal implants with titanium-sprayed surfaces. *Journal of Maxillofacial Surgery*. 1981;9:15-25. doi:10.1016/S0301-0503(81)80007-0
43. Sammons RL, Lumbikanonda N, Gross M, Cantzler P. Comparison of osteoblast spreading on microstructured dental implant surfaces and cell behaviour in an explant model of osseointegration: A scanning electron microscopic study. *Clinical Oral Implants Research*. 2005;16(6):657-666. doi:10.1111/j.1600-0501.2005.01168.x
44. Xue W, Liu X, Zheng X, Ding C. In vivo evaluation of plasma-sprayed titanium coating after alkali modification. *Biomaterials*. 2005;26(16):3029-3037. doi:10.1016/j.biomaterials.2004.09.003
45. Piattelli A, Scarano A, Corigliano M, Piattelli M. Effects of alkaline phosphatase on bone healing around plasma-sprayed titanium

- implants: a pilot study in rabbits. *Biomaterials*. 1996;17(14):1443-1449. doi:10.1016/0142-9612(96)87288-7
46. Piattelli A, Corigliano M, Scarano A, Costigliola G, Paolantonio M. Immediate Loading of Titanium Plasma-Sprayed Implants: An Histologic Analysis in Monkeys. *Journal of Periodontology*. 1998;69(3):321-327. doi:10.1902/jop.1998.69.3.321
 47. Scarano A, Iezzi G, Petrone G, Marinho VC, Corigliano M, Piattelli A. Immediate Postextraction Implants: A Histologic and Histometric Analysis in Monkeys. *Journal of Oral Implantology*. 2000;26(3):163-169. doi:10.1563/1548-1336(2000)026<0163:IP IAHA>2.3.CO;2
 48. Piattelli A, Corigliano M, Scarano A. Microscopical observations of the osseous responses in early loaded human titanium implants: a report of two cases. *Biomaterials*. 1996;17(13):1333-1337. doi:10.1016/S0142-9612(96)80011-1
 49. Piattelli A, Paolantonio M, Corigliano M, Scarano A. Immediate Loading of Titanium Plasma-Sprayed Screw-Shaped Implants in Man: A Clinical and Histological Report of Two Cases. *Journal of Periodontology*. 1997;68(6):591-597. doi:10.1902/jop.1997.68.6.591
 50. Piattelli A, Scarano A, Piattelli M, Bertolai R, Panzoni E. Histologic aspects of the bone and soft tissues surrounding three titanium non-submerged plasma-sprayed implants retrieved at autopsy: a case report. *J Periodontol*. 1997;68(7):694-700. doi:10.1902/jop.1997.68.7.694
 51. Piattelli A, Scarano A, Dalla Nora A, De Bona G, Favero GA. Microscopical features in retrieved human Branemark implants: a report of 19 cases. *Biomaterials*. 1998;19(7-9):643-649. doi:10.1016/S0142-9612(97)00158-0
 52. Piattelli A, Corigliano M, Scarano A, Quaranta M. Bone reactions to early occlusal loading of two-stage titanium plasma-sprayed implants: a pilot study in monkeys. *Int J Periodontics Restorative Dent*. 1997;17(2):162-169.
 53. Carr AB, Gerard DA, Larsen PE. Histomorphometric analysis of implant anchorage for 3 types of dental implants following 6 months of healing in baboon jaws. *Int J Oral Maxillofac Implants*. 2000;15(6):785-791.
 54. Franchi M, Bacchelli B, Martini D, et al. Early detachment of titanium particles from various different surfaces of endosseous dental implants. *Biomaterials*. 2004;25(12):2239-2246. doi:10.1016/j.biomaterials.2003.09.017
 55. YW H, HL C, LT L, KC T, DT B, YK W. Effects of non-thermal plasma on sandblasted titanium dental implants in beagle dogs. *Journal of the Chinese Medical Association : JCMA*. 2018;81(10):920-925.
 56. Canullo L, Tallarico M, Botticelli D, Alccayhuaman KAA, Martins Neto EC, Xavier SP. Hard and soft tissue changes around implants activated using plasma of argon: A histomorphometric study in dog. *Clin Oral Implants Res*. 2018;29(4):389-395. doi:10.1111/clr.13134
 57. Coelho PG, Bonfante EA, Pessoa RS, et al. Characterisation of five different implant surfaces and their effect on osseointegration: a study in dogs. *J Periodontol*. 2011;82(5):742-750. doi:10.1902/jop.2010.100520
 58. Novaes AB, Souza SLS, de Oliveira PT, Souza AMMS. Histomorphometric analysis of the bone-implant contact obtained with 4 different implant surface treatments placed side by side in the dog mandible. *Int J Oral Maxillofac Implants*. 2002;17(3):377-383.
 59. Qiao S, Cao H, Zhao X, et al. Ag-plasma modification enhances bone apposition around titanium dental implants: an animal study in Labrador dogs. *Int J Nanomedicine*. 2015;10:653-664. doi:10.2147/IJN.S73467
 60. Mostafa D, Aboushelib M. Bioactive-hybrid-zirconia implant surface for enhancing osseointegration: an in vivo study. *Int J Implant Dent*. 2018;4(1):20. doi:10.1186/s40729-018-0129-3
 61. Huang Z, Wang Z, Li C, Yin K, Hao D, Lan J. Application of Plasma Sprayed Zirconia Coating in Dental Implant: Study in Implant. *The Journal of Oral Implantology*. Published online January 5, 2018. doi:10.1563/aaid-joi-D-17-00124
 62. Metzler P, von Wilmsky C, Stadlinger B, et al. Nano-crystalline diamond-coated titanium dental implants - a histomorphometric study in adult domestic pigs. *J Craniomaxillofac Surg*. 2013;41(6):532-538. doi:10.1016/j.jcms.2012.11.020
 63. Ballo AM, Bjöörn D, Astrand M, Palmquist A, Lausmaa J, Thomsen P. Bone response to physical-vapour-deposited titanium dioxide coatings on titanium implants. *Clin Oral Implants Res*. 2013;24(9):1009-1017. doi:10.1111/j.1600-0501.2012.02509.x
 64. Park JY, Gemmell CH, Davies JE. Platelet interactions with titanium: modulation of platelet activity by surface topography. *Biomaterials*. 2001;22(19):2671-2682. doi:10.1016/S0142-9612(01)00009-6
 65. Ogawa T, Nishimura I. Different bone integration profiles of turned and acid-etched implants associated with modulated expression

- of extracellular matrix genes. *Int J Oral Maxillofac Implants*. 2003;18(2):200-210.
66. Degidi M, Petrone G, Iezzi G, Piattelli A. Bone Contact Around Acid-etched Implants: A Histological and Histomorphometrical Evaluation of Two Human-retrieved Implants. *Journal of Oral Implantology*. 2003;29(1):13-18. doi:10.1563/1548-1336(2003)029<0013:BCAAIA>2.3.CO;2
 67. Weng D, Hoffmeyer M, Hürzeler MB, Richter EJ. Osseotite ® vs. machined surface in poor bone quality: A study in dogs. *Clinical Oral Implants Research*. 2003;14(6):703-708. doi:10.1046/j.0905-7161.2003.00955.x
 68. Testori T, Szmukler-Moncler S, Francetti L, et al. Immediate loading of Osseotite implants: a case report and histologic analysis after 4 months of occlusal loading. *Int J Periodontics Restorative Dent*. 2001;21(5):451-459.
 69. Trisi P, Lazzara R, Rao W, Rebaudi A. Bone-implant contact and bone quality: evaluation of expected and actual bone contact on machined and osseotite implant surfaces. *Int J Periodontics Restorative Dent*. 2002;22(6):535-545.
 70. Cho S. The removal torque of titanium screw inserted in rabbit tibia treated by dual acid etching. *Biomaterials*. 2003;24(20):3611-3617. doi:10.1016/S0142-9612(03)00218-7
 71. Pontes AEF, de Toledo CT, Garcia VG, Ribeiro FS, Sakakura CE. Torque Analysis of a Triple Acid-Etched Titanium Implant Surface. *ScientificWorldJournal*. 2015;2015:819879. doi:10.1155/2015/819879
 72. Jinno Y, Jimbo R, Tovar N, Teixeira HS, Witek L, Coelho PG. In Vivo Evaluation of Dual Acid-Etched and Grit-Blasted/Acid-Etched Implants With Identical Macrogeometry in High-Density Bone. *Implant Dent*. 2017;26(6):815-819. doi:10.1097/ID.0000000000000672
 73. de Jesus RNR, Stavropoulos A, Oliveira MTF, Soares PBF, Moura CCG, Zanetta-Barbosa D. Histomorphometric evaluation of a dual acid-etched vs. a chemically modified hydrophilic dual acid-etched implant surface. An experimental study in dogs. *Clin Oral Implants Res*. 2017;28(5):551-557. doi:10.1111/clr.12833
 74. Bonfante EA, Granato R, Marin C, et al. Early bone healing and biomechanical fixation of dual acid-etched and as-machined implants with healing chambers: an experimental study in dogs. *Int J Oral Maxillofac Implants*. 2011;26(1):75-82.
 75. Halldin A, Jimbo R, Johansson CB, Gretzer C, Jacobsson M. Improved osseointegration and interlocking capacity with dual acid-treated implants: a rabbit study. *Clin Oral Implants Res*. 2016;27(1):22-30. doi:10.1111/clr.12507
 76. Fabbro MD, Taschieri S, Canciani E, et al. Osseointegration of Titanium Implants With Different Rough Surfaces: A Histologic and Histomorphometric Study in an Adult Minipig Model. *Implant Dent*. 2017;26(3):357-366. doi:10.1097/ID.0000000000000560
 77. Freitas GP, Lopes HB, Martins-Neto EC, de Oliveira PT, Beloti MM, Rosa AL. Effect of Surface Nanotopography on Bone Response to Titanium Implant. *J Oral Implantol*. 2016;42(3):240-247. doi:10.1563/aaid-joi-D-14-00254
 78. Yoo D, Marin C, Freitas G, et al. Surface characterisation and in vivo evaluation of dual Acid-etched and grit-blasted/acid-etched implants in sheep. *Implant Dent*. 2015;24(3):256-262. doi:10.1097/ID.0000000000000248
 79. Klokkevold PR, Johnson P, Dadgostari S, Davies JE, Caputo A, Nishimura RD. Early endosseous integration enhanced by dual acid etching of titanium: a torque removal study in the rabbit: Early endosseous integration enhanced by dual acid etching. *Clinical Oral Implants Research*. 2001;12(4):350-357. doi:10.1034/j.1600-0501.2001.012004350.x
 80. Galli C, Guizzardi S, Passeri G, et al. Comparison of Human Mandibular Osteoblasts Grown on Two Commercially Available Titanium Implant Surfaces. *Journal of Periodontology*. 2005;76(3):364-372. doi:10.1902/jop.2005.76.3.364
 81. Buser D, Nydegger T, Hirt HP, Cochran DL, Nolte LP. Removal torque values of titanium implants in the maxilla of miniature pigs. *Int J Oral Maxillofac Implants*. 1998;13(5):611-619.
 82. Abrahamsson I, Berglundh T, Linder E, Lang NP, Lindhe J. Early bone formation adjacent to rough and turned endosseous implant surfaces. An experimental study in the dog. *Clin Oral Implants Res*. 2004;15(4):381-392. doi:10.1111/j.1600-0501.2004.01082.x
 83. Cochran DL, Schenk RK, Lussi A, Higginbottom FL, Buser D. Bone response to unloaded and loaded titanium implants with a sandblasted and acid-etched surface: A histometric study in the canine mandible. *J Biomed Mater Res*. 1998;40(1):1-11. doi:10.1002/(SICI)1097-4636(199804)40:1<1::AID-JBM1>3.0.CO;2-Q
 84. Buser D, Schenk RK, Steinemann S, Fiorellini JP, Fox CH, Stich H. Influence of surface characteristics on bone integration of titanium implants. A histomorphometric study in miniature pigs. *J Biomed Mater Res*. 1991;25(7):889-902. doi:10.1002/

jbm.820250708

85. Wilke HJ. The influence of various titanium surfaces on the interface shear strength between implants and bone. *Clinical implant materials Adv Biomater*. 1990;9:309-314.
86. Hayakawa T, Kiba H, Yasuda S, Yamamoto H, Nemoto K. A histologic and histomorphometric evaluation of two types of retrieved human titanium implants. *Int J Periodontics Restorative Dent*. 2002;22(2):164-171.
87. Sakakura CE, Nociti FH, Mello GPS, de Mello EDA, de Rezende MLR. Histomorphometric Evaluation of a Threaded, Sandblasted, Acid-Etched Implant Retrieved From a Human Lower Jaw: A Case Report. *Implant Dentistry*. 2005;14(3):289-293. doi:10.1097/01.id.0000173641.57293.e4
88. Cochran DL, Buser D, Ten Bruggenkate CM, et al. The use of reduced healing times on ITI ® implants with a sandblasted and acid-etched (SLA) surface:: Early results from clinical trials on ITI ® SLA implants. *Clinical Oral Implants Research*. 2002;13(2):144-153. doi:10.1034/j.1600-0501.2002.130204.x
89. Guizzardi S, Galli C, Martini D, et al. Different Titanium Surface Treatment Influences Human Mandibular Osteoblast Response. *Journal of Periodontology*. 2004;75(2):273-282. doi:10.1902/jop.2004.75.2.273
90. Dundar S, Yaman F, Bozoglan A, et al. Comparison of Osseointegration of Five Different Surfaced Titanium Implants. *J Craniofac Surg*. 2018;29(7):1991-1995. doi:10.1097/SCS.00000000000004572
91. Chiang HJ, Hsu HJ, Peng PW, et al. Early bone response to machined, sandblasting acid etching (SLA) and novel surface-functionalisation (SLAffinity) titanium implants: characterisation, biomechanical analysis and histological evaluation in pigs. *J Biomed Mater Res A*. 2016;104(2):397-405. doi:10.1002/jbm.a.35577
92. Ernst S, Stübinger S, Schüpbach P, et al. Comparison of two dental implant surface modifications on implants with same macrodesign: an experimental study in the pelvic sheep model. *Clin Oral Implants Res*. 2015;26(8):898-908. doi:10.1111/clr.12411
93. Calvo-Guirado JL, Satorres M, Negri B, et al. Biomechanical and histological evaluation of four different titanium implant surface modifications: an experimental study in the rabbit tibia. *Clin Oral Investig*. 2014;18(5):1495-1505. doi:10.1007/s00784-013-1120-2
94. Lai HC, Zhuang LF, Zhang ZY, Wieland M, Liu X. Bone apposition around two different sandblasted, large-grit and acid-etched implant surfaces at sites with coronal circumferential defects: an experimental study in dogs. *Clin Oral Implants Res*. 2009;20(3):247-253. doi:10.1111/j.1600-0501.2008.01651.x
95. Al-Nawas B, Groetz KA, Goetz H, Duschner H, Wagner W. Comparative histomorphometry and resonance frequency analysis of implants with moderately rough surfaces in a loaded animal model. *Clin Oral Implants Res*. 2008;19(1):1-8. doi:10.1111/j.1600-0501.2007.01396.x
96. Perrin D, Szmukler-Moncler S, Echikou C, Pointaire P, Bernard JP. Bone response to alteration of surface topography and surface composition of sandblasted and acid etched (SLA) implants. *Clin Oral Implants Res*. 2002;13(5):465-469. doi:10.1034/j.1600-0501.2002.130504.x
97. Marinho VC, Celletti R, Bracchetti G, Petrone G, Minkin C, Piattelli A. Sandblasted and acid-etched dental implants: a histologic study in rats. *Int J Oral Maxillofac Implants*. 2003;18(1):75-81.
98. Buser D, Nydegger T, Oxland T, et al. Interface shear strength of titanium implants with a sandblasted and acid-etched surface: A biomechanical study in the maxilla of miniature pigs. *J Biomed Mater Res*. 1999;45(2):75-83. doi:10.1002/(SICI)1097-4636(199905)45:2<75::AID-JBM1>3.0.CO;2-P
99. Sul YT, Johansson CB, Petronis S, et al. Characteristics of the surface oxides on turned and electrochemically oxidised pure titanium implants up to dielectric breakdown: *Biomaterials*. 2002;23(2):491-501. doi:10.1016/S0142-9612(01)00131-4
100. Xavier SP, Ikuno KE, Tavares MG. Enhanced bone apposition to Brazilian microrough titanium surfaces. *Brazilian dental journal*. 2010;21:18-23.
101. Zhu X, Chen J, Scheideler L, Reichl R, Geis-Gerstorf J. Effects of topography and composition of titanium surface oxides on osteoblast responses. *Biomaterials*. 2004;25(18):4087-4103. doi:10.1016/j.biomaterials.2003.11.011
102. Son W woo, Zhu X, Shin H in, Ong JL, Kim K han. In vivo histological response to anodised and anodised/hydrothermally treated titanium implants. *J Biomed Mater Res*. 2003;66B(2):520-525. doi:10.1002/jbm.b.10042

103. Choi JW, Heo SJ, Koak JY, et al. Biological responses of anodised titanium implants under different current voltages. *J Oral Rehabil.* 2006;33(12):889-897. doi:10.1111/j.1365-2842.2006.01669.x
104. Yamagami A, Yoshihara Y, Suwa F. Mechanical and histologic examination of titanium alloy material treated by sandblasting and anodic oxidisation. *Int J Oral Maxillofac Implants.* 2005;20(1):48-53.
105. Kang BS, Sul YT, Johansson CB, Oh SJ, Lee HJ, Albrektsson T. The effect of calcium ion concentration on the bone response to oxidised titanium implants. *Clin Oral Implants Res.* 2012;23(6):690-697. doi:10.1111/j.1600-0501.2011.02177.x
106. Knobloch L, Larsen PA, Rashid B, Carr AB. Six-month performance of implants with oxidised and machined surfaces restored at 2, 4, and 6 weeks postimplantation in adult beagle dogs. *Int J Oral Maxillofac Implants.* 2004;19(3):350-356.
107. Lee HJ, Yang IH, Kim SK, Yeo IS, Kwon TK. In vivo comparison between the effects of chemically modified hydrophilic and anodically oxidised titanium surfaces on initial bone healing. *J Periodontal Implant Sci.* 2015;45(3):94-100. doi:10.5051/jpis.2015.45.3.94
108. Park IP, Kang TJ, Heo SJ, et al. Investigation of anodised titanium implants coated with triterpenoids extracted from black cohosh: an animal study. *J Adv Prosthodont.* 2014;6(1):14-21. doi:10.4047/jap.2014.6.1.14
109. Sul YT, Johansson CB, Kang Y, Jeon DG, Albrektsson T. Bone reactions to oxidised titanium implants with electrochemical anion sulphuric acid and phosphoric acid incorporation. *Clin Implant Dent Relat Res.* 2002;4(2):78-87. doi:10.1111/j.1708-8208.2002.tb00156.x
110. Li X, Xu H, Zhao B, Jiang S. Accelerated and enhanced osteointegration of MAO-treated implants: histological and histomorphometric evaluation in a rabbit model. *Int J Oral Sci.* 2018;10(2):11. doi:10.1038/s41368-018-0008-z
111. Ma W, Wei JH, Li YZ, et al. Histological evaluation and surface componential analysis of modified micro-arc oxidation-treated titanium implants. *J Biomed Mater Res B Appl Biomater.* 2008;86(1):162-169. doi:10.1002/jbm.b.31002
112. Sul YT, Johansson CB, Albrektsson T. Oxidized titanium screws coated with calcium ions and their performance in rabbit bone. *Int J Oral Maxillofac Implants.* 2002;17(5):625-634.
113. Ivanoff CJ, Widmark G, Johansson C, Wennerberg A. Histologic evaluation of bone response to oxidised and turned titanium micro-implants in human jawbone. *Int J Oral Maxillofac Implants.* 2003;18(3):341-348.
114. Xie J, Baumann MJ, McCabe LR. Osteoblasts respond to hydroxyapatite surfaces with immediate changes in gene expression. *J Biomed Mater Res.* 2004;71A(1):108-117. doi:10.1002/jbm.a.30140
115. Uehara T, Takaoka K, Ito K. Histological evidence of osseointegration in human retrieved fractured hydroxyapatite-coated screw-type implants: a case report. *Clin Oral Implants Res.* 2004;15(5):540-545. doi:10.1111/j.1600-0501.2004.01031.x
116. Morris HF, Ochi S. Hydroxyapatite-coated implants: A case for their use. *Journal of Oral and Maxillofacial Surgery.* 1998;56(11):1303-1311. doi:10.1016/S0278-2391(98)90615-2
117. Lee JJ, Rouhfar L, Beirne OR. Survival of hydroxyapatite-coated implants: A meta-analytic review. *Journal of Oral and Maxillofacial Surgery.* 2000;58(12):1372-1379. doi:10.1053/joms.2000.18269
118. Trisi P, Keith DJ, Rocco S. Human histologic and histomorphometric analyses of hydroxyapatite-coated implants after 10 years of function: a case report. *Int J Oral Maxillofac Implants.* 2005;20(1):124-130.
119. Piattelli A, Cosci F, Scarano A, Trisi P. Localised chronic suppurative bone infection as a sequel of peri-implantitis in a hydroxyapatite-coated dental implant. *Biomaterials.* 1995;16(12):917-920. doi:10.1016/0142-9612(95)93116-U
120. Sun L, Berndt CC, Gross KA, Kucuk A. Material fundamentals and clinical performance of plasma-sprayed hydroxyapatite coatings: A review. *J Biomed Mater Res.* 2001;58(5):570-592. doi:10.1002/jbm.1056
121. Ong JL, Carnes DL, Bessho K. Evaluation of titanium plasma-sprayed and plasma-sprayed hydroxyapatite implants in vivo. *Biomaterials.* 2004;25(19):4601-4606. doi:10.1016/j.biomaterials.2003.11.053
122. Piattelli M, Scarano A, Paolantonio M, Iezzi G, Petrone G, Piattelli A. Bone Response to Machined and Resorbable Blast Material Titanium Implants: An Experimental Study in Rabbits. *Journal of Oral Implantology.* 2002;28(1):2-8. doi:10.1563/1548-1336(2002)028<0002:BRTMAR>2.3.CO;2
123. Jung YC, Han CH, Lee IS, Kim HE. Effects of ion beam-assisted deposition of hydroxyapatite on the osseointegration of

- endosseous implants in rabbit tibiae. *Int J Oral Maxillofac Implants*. 2001;16(6):809-818.
124. Lee IS, Kim DH, Kim HE, Jung YC, Han CH. Biological performance of calcium phosphate films formed on commercially pure Ti by electron-beam evaporation. *Biomaterials*. 2002;23(2):609-615. doi:10.1016/S0142-9612(01)00147-8
125. Svanborg LM, Hoffman M, Andersson M, Currie F, Kjellin P, Wennerberg A. The effect of hydroxyapatite nanocrystals on early bone formation surrounding dental implants. *Int J Oral Maxillofac Surg*. 2011;40(3):308-315. doi:10.1016/j.ijom.2010.10.010
126. Scarano A, Piattelli A, Quaranta A, Lorusso F. Bone Response to Two Dental Implants with Different Sandblasted/Acid-Etched Implant Surfaces: A Histological and Histomorphometrical Study in Rabbits. *BioMed Research International*. 2017;2017:8724951. doi:10.1155/2017/8724951
127. Chan YH, Lew WZ, Lu E, et al. An evaluation of the biocompatibility and osseointegration of novel glass fiber reinforced composite implants: In vitro and in vivo studies. *Dent Mater*. 2018;34(3):470-485. doi:10.1016/j.dental.2017.12.001
128. Trisi P, Berardini M, Falco A, Sandrini E, Vulpiani MP. A New Highly Hydrophilic Electrochemical Implant Titanium Surface: A Histological and Biomechanical In Vivo Study. *Implant Dentistry*. 2017;26(3):429-437. doi:10.1097/ID.0000000000000605
129. Cardoso MV, de Rycker J, Chaudhari A, et al. Titanium implant functionalisation with phosphate-containing polymers may favour in vivo osseointegration. *J Clin Periodontol*. 2017;44(9):950-960. doi:10.1111/jcpe.12736
130. Liu Y, Zhou Y, Jiang T, Liang YD, Zhang Z, Wang YN. Evaluation of the osseointegration of dental implants coated with calcium carbonate: an animal study. *Int J Oral Sci*. 2017;9(3):133-138. doi:10.1038/ijos.2017.13
131. Kalemaj Z, Scarano A, Valbonetti L, Rapone B, Grassi FR. Bone Response to Four Dental Implants with Different Surface Topographies: A Histologic and Histometric Study in Minipigs. *The International Journal of Periodontics & Restorative Dentistry*. 2016;36(5):745-754. doi:10.11607/prd.2719
132. Song WW, Heo JH, Lee JH, Park YM, Kim YD. Osseointegration of magnesium-incorporated sandblasted acid-etched implant in the dog mandible: Resonance frequency measurements and histomorphometric analysis. *Tissue Eng Regen Med*. 2016;13(2):191-199. doi:10.1007/s13770-016-9126-x
133. Mistry S, Roy S, Jyoti Maitra N, et al. Safety and efficacy of additive and subtractive surface modification of Ti6Al4V endosseous implant in goat bone. *J Mech Behav Biomed Mater*. 2016;57:69-87. doi:10.1016/j.jmbbm.2015.11.019
134. Armencea G, Berce C, Rotaru H, et al. Micro-CT and histological analysis of Ti6Al7Nb custom made implants with hydroxyapatite and SiO₂-TiO₂ coatings in a rabbit model. *Clujul Med*. 2015;88(3):408-414. doi:10.15386/cjmed-479
135. Im JH, Kim SG, Oh JS, Lim SC. A Comparative Study of Stability After the Installation of 2 Different Surface Types of Implants in the Maxillae of Dogs. *Implant Dent*. 2015;24(5):586-591. doi:10.1097/ID.0000000000000292
136. Melin Svanborg L, Meirelles L, Franke Stenport V, et al. Evaluation of bone healing on sandblasted and acid etched implants coated with nanocrystalline hydroxyapatite: an in vivo study in rabbit femur. *Int J Dent*. 2014;2014:197581. doi:10.1155/2014/197581
137. Bryington MS, Hayashi M, Kozai Y, et al. The influence of nano hydroxyapatite coating on osseointegration after extended healing periods. *Dent Mater*. 2013;29(5):514-520. doi:10.1016/j.dental.2013.02.004
138. Eom TG, Jeon GR, Jeong CM, et al. Experimental study of bone response to hydroxyapatite coating implants: bone-implant contact and removal torque test. *Oral Surg Oral Med Oral Pathol Oral Radiol*. 2012;114(4):411-418. doi:10.1016/j.oooo.2011.10.036
139. Choi JY, Jung UW, Kim CS, Jung SM, Lee IS, Choi SH. Influence of nanocoated calcium phosphate on two different types of implant surfaces in different bone environment: an animal study. *Clin Oral Implants Res*. 2013;24(9):1018-1022. doi:10.1111/j.1600-0501.2012.02492.x
140. Poulos NM, Rodriguez NA, Lee J, et al. Evaluation of a novel calcium phosphate-coated titanium porous oxide implant surface: a study in rabbits. *Int J Oral Maxillofac Implants*. 2011;26(4):731-738.
141. Mano T, Ishikawa K, Harada K, Umeda H, Ueyama Y. Comparison of apatite-coated titanium prepared by blast coating and flame spray methods--evaluation using simulated body fluid and initial histological study. *Dent Mater J*. 2011;30(4):431-437. doi:10.4012/dmj.2010-162
142. Jimbo R, Sotres J, Johansson C, Breiding K, Currie F, Wennerberg A. The biological response to three different nanostructures applied on smooth implant surfaces. *Clin Oral Implants Res*. 2012;23(6):706-712. doi:10.1111/j.1600-0501.2011.02182.x

143. Coelho PG, Granato R, Marin C, et al. Effect of Si addition on Ca- and P-impregnated implant surfaces with nanometer-scale roughness: an experimental study in dogs. *Clin Oral Implants Res.* 2012;23(3):373-378. doi:10.1111/j.1600-0501.2010.02150.x
144. Suzuki M, Guimaraes MVM, Marin C, et al. Histomorphologic and bone-to-implant contact evaluation of dual acid-etched and bioceramic grit-blasted implant surfaces: an experimental study in dogs. *J Oral Maxillofac Surg.* 2010;68(8):1877-1883. doi:10.1016/j.joms.2009.09.050
145. Junker R, Manders PJD, Wolke J, Borisov Y, Jansen JA. Bone-supportive behavior of microplasma-sprayed CaP-coated implants: mechanical and histological outcome in the goat. *Clin Oral Implants Res.* 2010;21(2):189-200. doi:10.1111/j.1600-0501.2009.01819.x
146. Lin A, Wang CJ, Kelly J, Gubbi P, Nishimura I. The role of titanium implant surface modification with hydroxyapatite nanoparticles in progressive early bone-implant fixation in vivo. *Int J Oral Maxillofac Implants.* 2009;24(5):808-816.
147. Barros RRM, Novaes AB, Papalexiou V, et al. Effect of biofunctionalised implant surface on osseointegration: a histomorphometric study in dogs. *Braz Dent J.* 2009;20(2):91-98. doi:10.1590/s0103-64402009000200001
148. Suzuki M, Guimaraes MVM, Marin C, Granato R, Gil JN, Coelho PG. Histomorphometric evaluation of alumina-blasted/acid-etched and thin ion beam-deposited bioceramic surfaces: an experimental study in dogs. *J Oral Maxillofac Surg.* 2009;67(3):602-607. doi:10.1016/j.joms.2008.08.021
149. Yang G li, He F ming, Hu J an, Wang X xiang, Zhao S fang. Effects of biomimetically and electrochemically deposited nano-hydroxyapatite coatings on osseointegration of porous titanium implants. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2009;107(6):782-789. doi:10.1016/j.tripleo.2008.12.023
150. Granato R, Marin C, Suzuki M, Gil JN, Janal MN, Coelho PG. Biomechanical and histomorphometric evaluation of a thin ion beam bioceramic deposition on plateau root form implants: an experimental study in dogs. *J Biomed Mater Res B Appl Biomater.* 2009;90(1):396-403. doi:10.1002/jbm.b.31298
151. Le Guehennec L, Goyenvalle E, Lopez-Heredia MA, Weiss P, Amouriq Y, Layrolle P. Histomorphometric analysis of the osseointegration of four different implant surfaces in the femoral epiphyses of rabbits. *Clin Oral Implants Res.* 2008;19(11):1103-1110. doi:10.1111/j.1600-0501.2008.01547.x
152. Marin C, Granato R, Suzuki M, Gil JN, Piattelli A, Coelho PG. Removal torque and histomorphometric evaluation of bioceramic grit-blasted/acid-etched and dual acid-etched implant surfaces: an experimental study in dogs. *J Periodontol.* 2008;79(10):1942-1949. doi:10.1902/jop.2008.080106
153. Coelho PG, Cardaropoli G, Suzuki M, Lemons JE. Histomorphometric evaluation of a nanothickness bioceramic deposition on endosseous implants: a study in dogs. *Clin Implant Dent Relat Res.* 2009;11(4):292-302. doi:10.1111/j.1708-8208.2008.00122.x
154. Alzubaydi TL, Alameer SS, Ismaeel T, Alhijazi AY, Geetha M. In vivo studies of the ceramic coated titanium alloy for enhanced osseointegration in dental applications. *J Mater Sci Mater Med.* 2009;20 Suppl 1:S35-42. doi:10.1007/s10856-008-3479-1
155. Franco R de L, Chiesa R, de Oliveira PT, Beloti MM, Rosa AL. Bone response to a Ca- and P-enriched titanium surface obtained by anodisation. *Braz Dent J.* 2008;19(1):15-20. doi:10.1590/s0103-64402008000100003
156. Yeo IS, Han JS, Yang JH. Biomechanical and histomorphometric study of dental implants with different surface characteristics. *Journal of Biomedical Materials Research Part B: Applied Biomaterials: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials.* 2008;87(2):303-311.
157. Zagury R, Harari ND, Conz MB, Soares G de A, Vidigal GM. Histomorphometric analyses of bone interface with titanium-aluminum-vanadium and hydroxyapatite-coated implants by biomimetic process. *Implant Dent.* 2007;16(3):290-296. doi:10.1097/ID.0b013e3180e9d9ed
158. Siebers MC, Wolke JGC, Walboomers XF, Leeuwenburgh SCG, Jansen JA. In vivo evaluation of the trabecular bone behavior to porous electrostatic spray deposition-derived calcium phosphate coatings. *Clin Oral Implants Res.* 2007;18(3):354-361. doi:10.1111/j.1600-0501.2006.01314.x
159. Hayakawa T, Takahashi K, Yoshinari M, et al. Trabecular bone response to titanium implants with a thin carbonate-containing

- apatite coating applied using the molecular precursor method. *Int J Oral Maxillofac Implants*. 2006;21(6):851-858.
160. Manders PJD, Wolke JGC, Jansen JA. Bone response adjacent to calcium phosphate electrostatic spray deposition coated implants: an experimental study in goats. *Clin Oral Implants Res*. 2006;17(5):548-553. doi:10.1111/j.1600-0501.2006.01263.x
161. Xiropaidis AV, Qahash M, Lim WH, et al. Bone-implant contact at calcium phosphate-coated and porous titanium oxide (TiUnite)-modified oral implants. *Clin Oral Implants Res*. 2005;16(5):532-539. doi:10.1111/j.1600-0501.2005.01126.x
162. Schopper C, Moser D, Goriwoda W, et al. The effect of three different calcium phosphate implant coatings on bone deposition and coating resorption: a long-term histological study in sheep. *Clin Oral Implants Res*. 2005;16(3):357-368. doi:10.1111/j.1600-0501.2004.01080.x
163. Hayakawa T, Yoshinari M, Kiba H, Yamamoto H, Nemoto K, Jansen JA. Trabecular bone response to surface roughened and calcium phosphate (Ca-P) coated titanium implants. *Biomaterials*. 2002;23(4):1025-1031. doi:10.1016/S0142-9612(01)00214-9
164. Strnad Z, Strnad J, Povýsil C, Urban K. Effect of plasma-sprayed hydroxyapatite coating on the osteoconductivity of commercially pure titanium implants. *Int J Oral Maxillofac Implants*. 2000;15(4):483-490.
165. Hulshoff JE, van Dijk K, van der Waerden JP, Wolke JG, Kalk W, Jansen JA. Evaluation of plasma-spray and magnetron-sputter Ca-P-coated implants: an in vivo experiment using rabbits. *J Biomed Mater Res*. 1996;31(3):329-337. doi:10.1002/(SICI)1097-4636(199607)31:3<329::AID-JBM6>3.0.CO;2-O
166. Yoshinari M, Klinge B, Dérand T. The biocompatibility (cell culture and histologic study) of hydroxy-apatite-coated implants created by ion beam dynamic mixing. *Clin Oral Implants Res*. 1996;7(2):96-100. doi:10.1034/j.1600-0501.1996.070202.x
167. Caulier H, van der Waerden JP, Paquay YC, et al. Effect of calcium phosphate (Ca-P) coatings on trabecular bone response: a histological study. *J Biomed Mater Res*. 1995;29(9):1061-1069. doi:10.1002/jbm.820290906
168. OG B, C C, MA A, S AH, ML E, MC A. Osseointegration of Ti6Al4V dental implants modified by thermal oxidation in osteoporotic rabbits. *International journal of implant dentistry*. 2016;2(1):18.
169. Ozeki K, Okuyama Y, Fukui Y, Aoki H. Bone response to titanium implants coated with thin sputtered HA film subject to hydrothermal treatment and implanted in the canine mandible. *Biomed Mater Eng*. 2006;16(4):243-251.
170. García-Alonso MC, Saldaña L, Vallés G, et al. In vitro corrosion behaviour and osteoblast response of thermally oxidised Ti6Al4V alloy. *Biomaterials*. 2003;24(1):19-26. doi:10.1016/s0142-9612(02)00237-5
171. Scarano A, Crocetta E, Quaranta A, Lorusso F. Influence of the thermal treatment to address a better osseointegration of Ti6Al4V Dental Implants: Histological and histomorphometrical study in a rabbit model. *BioMed Research International*. 2018;2018. doi:10.1155/2018/2349698
172. Herrero-Climent M, Romero Ruiza MM, Calvo PL, Santos JVR, Perez RA, Gil Mur FJ. Effectiveness of a new dental implant bioactive surface: histological and histomorphometric comparative study in minipigs. *Clin Oral Investig*. 2018;22(3):1423-1432. doi:10.1007/s00784-017-2223-y
173. Qamheya AHA, Arisan V, Mutlu Z, et al. Thermal oxidation and hydrofluoric acid treatment on the sandblasted implant surface: A histologic histomorphometric and biomechanical study. *Clin Oral Implants Res*. 2018;29(7):741-755. doi:10.1111/clr.13285
174. Kim NS, Vang MS, Yang HS, Park SW, Park HO, Lim HP. Comparison of stability in titanium implants with different surface topographies in dogs. *J Adv Prosthodont*. 2009;1(1):47-55. doi:10.4047/jap.2009.1.1.47
175. Piconi C, Maccauro G. Zirconia as a ceramic biomaterial. *Biomaterials*. 1999;20(1):1-25.
176. Han JM, Hong G, Lin H, et al. Biomechanical and histological evaluation of the osseointegration capacity of two types of zirconia implant. *Int J Nanomedicine*. 2016;11:6507-6516. doi:10.2147/IJN.S119519
177. Martins R, Cestari TM, Arantes RVN, et al. Osseointegration of zirconia and titanium implants in a rabbit tibiae model evaluated by microtomography, histomorphometry and fluorochrome labeling analyses. *J Periodontol Res*. 2018;53(2):210-221. doi:10.1111/jre.12508
178. Mihatovic I, Golubovic V, Becker J, Schwarz F. Bone tissue response to experimental zirconia implants. *Clinical oral investigations*. 2017;21(2):523-532.
179. Jimbo R, Naito Y, Galli S, Berner S, Dard M, Wennerberg A. Biomechanical and Histomorphometrical Evaluation of TiZr Alloy

- Implants: An in vivo Study in the Rabbit. *Clin Implant Dent Relat Res*. 2015;17 Suppl 2:e670-678. doi:10.1111/cid.12305
180. Gredes T, Kubasiewicz-Ross P, Gedrange T, Dominiak M, Kunert-Keil C. Comparison of surface modified zirconia implants with commercially available zirconium and titanium implants: a histological study in pigs. *Implant Dent*. 2014;23(4):502-507. doi:10.1097/ID.000000000000110
181. Saulacic N, Erdösi R, Bosshardt DD, Gruber R, Buser D. Acid and alkaline etching of sandblasted zirconia implants: a histomorphometric study in miniature pigs. *Clin Implant Dent Relat Res*. 2014;16(3):313-322. doi:10.1111/cid.12070
182. Hoffmann O, Angelov N, Zafiroopoulos GG, Andreana S. Osseointegration of zirconia implants with different surface characteristics: an evaluation in rabbits. *Int J Oral Maxillofac Implants*. 2012;27(2):352-358.
183. Lee J, Sieweke JH, Rodriguez NA, et al. Evaluation of nano-technology-modified zirconia oral implants: a study in rabbits. *J Clin Periodontol*. 2009;36(7):610-617. doi:10.1111/j.1600-051X.2009.01423.x
184. Kohal RJ, Wolkewitz M, Hinze M, Han JS, Bächle M, Butz F. Biomechanical and histological behavior of zirconia implants: an experiment in the rat. *Clin Oral Implants Res*. 2009;20(4):333-339. doi:10.1111/j.1600-0501.2008.01656.x
185. Depprich R, Zipprich H, Ommerborn M, et al. Osseointegration of zirconia implants compared with titanium: an in vivo study. *Head Face Med*. 2008;4:30. doi:10.1186/1746-160X-4-30
186. Langhoff JD, Voelter K, Scharnweber D, et al. Comparison of chemically and pharmaceutically modified titanium and zirconia implant surfaces in dentistry: a study in sheep. *Int J Oral Maxillofac Surg*. 2008;37(12):1125-1132. doi:10.1016/j.ijom.2008.09.008
187. Gahlert M, Gudehus T, Eichhorn S, Steinhäuser E, Kniha H, Erhardt W. Biomechanical and histomorphometric comparison between zirconia implants with varying surface textures and a titanium implant in the maxilla of miniature pigs. *Clin Oral Implants Res*. 2007;18(5):662-668. doi:10.1111/j.1600-0501.2007.01401.x
188. Sennerby L, Dasmah A, Larsson B, Iverhed M. Bone tissue responses to surface-modified zirconia implants: A histomorphometric and removal torque study in the rabbit. *Clin Implant Dent Relat Res*. 2005;7 Suppl 1:S13-20. doi:10.1111/j.1708-8208.2005.tb00070.x
189. Dubruille JH, Viguier E, Le Naour G, Dubruille MT, Auriol M, Le Charpentier Y. Evaluation of combinations of titanium, zirconia, and alumina implants with 2 bone fillers in the dog. *Int J Oral Maxillofac Implants*. 1999;14(2):271-277.
190. Kohal RJ, Weng D, Bächle M, Strub JR. Loaded Custom-Made Zirconia and Titanium Implants Show Similar Osseointegration: An Animal Experiment. *Journal of Periodontology*. 2004;75(9):1262-1268. doi:10.1902/jop.2004.75.9.1262
191. Albrektsson T, Hansson H, Ivarsson B. Interface analysis of titanium and zirconium bone implants. *Biomaterials*. 1985;6(2):97-101. doi:10.1016/0142-9612(85)90070-5
192. Thomsen P, Larsson C, Ericson LE, Sennerby L, Lausmaa J, Kasemo B. Structure of the interface between rabbit cortical bone and implants of gold, zirconium and titanium. *Journal of Materials Science: Materials in Medicine*. 1997;8(11):653-665.
193. Ichikawa Y, Akagawa Y, Nikai H, Tsuru H. Tissue compatibility and stability of a new zirconia ceramic in vivo. *J Prosthet Dent*. 1992;68(2):322-326. doi:10.1016/0022-3913(92)90338-b
194. Akagawa Y, Ichikawa Y, Nikai H, Tsuru H. Interface histology of unloaded and early loaded partially stabilised zirconia endosseous implant in initial bone healing. *J Prosthet Dent*. 1993;69(6):599-604. doi:10.1016/0022-3913(93)90289-z
195. Covacci V, Bruzzese N, Maccauro G, et al. In vitro evaluation of the mutagenic and carcinogenic power of high purity zirconia ceramic. *Biomaterials*. 1999;20(4):371-376. doi:10.1016/S0142-9612(98)00182-3
196. Schultze-Mosgau S, Schliephake H, Radespiel-Tröger M, Neukam FW. Osseointegration of endodontic endosseous cones Zirconium oxide vs titanium. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*. 2000;89(1):91-98. doi:10.1016/S1079-2104(00)80022-0
197. Josset Y, Oum'Hamed Z, Zarrinpour A, Lorenzato M, Adnet JJ, Laurent-Maquin D. In vitro reactions of human osteoblasts in culture with zirconia and alumina ceramics. *J Biomed Mater Res*. 1999;47(4):481-493. doi:10.1002/(SICI)1097-4636(19991215)47:4<481::AID-JBM4>3.0.CO;2-Y
198. Scarano A, Di Carlo F, Quaranta M, Piattelli A. Bone response to zirconia ceramic implants: an experimental study in rabbits. *The Journal of Oral Implantology*. 2003;29(1):8-12. doi:10.1563/1548-1336(2003)029<0008:BRTZCI>2.3.CO;2
199. Akagawa Y, Hosokawa R, Sato Y, Kamayama K. Comparison between freestanding and tooth-connected partially stabilised

- zirconia implants after two years' function in monkeys: a clinical and histologic study. *J Prosthet Dent*. 1998;80(5):551-558. doi:10.1016/s0022-3913(98)70031-9
200. Okamoto K, Kido H, Sato A, Watazu A, Matsuura M. Ultrastructure of the interface between titanium and surrounding tissue in rat tibiae--a comparison study on titanium-coated and -uncoated plastic implants. *Clin Implant Dent Relat Res*. 2007;9(2):100-111. doi:10.1111/j.1708-8208.2007.00032.x
201. Mehl C, Kern M, Neumann F, Bähr T, Wiltfang J, Gassling V. Effect of ultraviolet photofunctionalization of dental titanium implants on osseointegration. *J Zhejiang Univ Sci B*. 2018;19(7):525-534. doi:10.1631/jzus.B1600505
202. Yamauchi R, Itabashi T, Wada K, Tanaka T, Kumagai G, Ishibashi Y. Photofunctionalised Ti6Al4V implants enhance early phase osseointegration. *Bone Joint Res*. 2017;6(5):331-336. doi:10.1302/2046-3758.65.BJR-2016-0221.R1
203. Shen J, Liu J, Chen X, Wang X, He F, Wang H. The In Vivo Bone Response of Ultraviolet-Irradiated Titanium Implants Modified with Spontaneously Formed Nanostructures: An Experimental Study in Rabbits. *Int J Oral Maxillofac Implants*. 2016;31(4):776-784. doi:10.11607/jomi.4309
204. Hirakawa Y, Jimbo R, Shibata Y, Watanabe I, Wennerberg A, Sawase T. Accelerated bone formation on photo-induced hydrophilic titanium implants: an experimental study in the dog mandible. *Clin Oral Implants Res*. 2013;24 Suppl A100:139-144. doi:10.1111/j.1600-0501.2011.02401.x
205. Fiorellini JP, Glindmann S, Salcedo J, Weber HP, Park CJ, Sarmiento HL. The Effect of Osteopontin and an Osteopontin-Derived Synthetic Peptide Coating on Osseointegration of Implants in a Canine Model. *Int J Periodontics Restorative Dent*. 2016;36(6):e88-e94. doi:10.11607/prd.2830
206. Johansson CB, Han CH, Wennerberg A, Albrektsson T. A quantitative comparison of machined commercially pure titanium and titanium-aluminum-vanadium implants in rabbit bone. *International Journal of Oral & Maxillofacial Implants*. 1998;13(3).
207. Carr AB, Larsen PE, Gerard DA. Histomorphometric comparison of implant anchorage for two types of dental implants after 3 and 6 months' healing in baboon jaws. *The Journal of Prosthetic Dentistry*. 2001;85(3):276-280. doi:10.1067/mpr.2001.114821
208. Lausmaa J, Mattsson L, Rolander U, Kasemo B. Chemical Composition and Morphology of Titanium Surface Oxides. *MRS Proc*. 1985;55:351. doi:10.1557/PROC-55-351
209. Arys A, Philippart C, Douvrou N, He Y, Le QT, Pireaux JJ. Analysis of titanium dental implants after failure of osseointegration: Combined histological, electron microscopy, and X-ray photoelectron spectroscopy approach. *J Biomed Mater Res*. 1998;43(3):300-312. doi:10.1002/(SICI)1097-4636(199823)43:3<300::AID-JBM11>3.0.CO;2-J
210. Zitter H, Plenck H. The electrochemical behavior of metallic implant materials as an indicator of their biocompatibility. *J Biomed Mater Res*. 1987;21(7):881-896. doi:10.1002/jbm.820210705
211. Ducheyne P. Titanium and calcium phosphate ceramic dental implants, surfaces, coatings and interfaces. *The Journal of Oral Implantology*. 1988;14(3):325-340.
212. Larsson C, Thomsen P, Aronsson BO, et al. Bone response to surface-modified titanium implants: studies on the early tissue response to machined and electropolished implants with different oxide thicknesses. *Biomaterials*. 1996;17(6):605-616. doi:10.1016/0142-9612(96)88711-4
213. Olefjord I, Hansson S. Surface analysis of four dental implant systems. *International Journal of Oral & Maxillofacial Implants*. 1993;8(1).
214. Lim YJ, Oshida Y, Andres CJ, Barco MT. Surface characterisations of variously treated titanium materials. *International Journal of Oral & Maxillofacial Implants*. 2001;16(3).
215. Sul YT, Johansson CB, Röser K, Albrektsson T. Qualitative and quantitative observations of bone tissue reactions to anodised implants. *Biomaterials*. 2002;23(8):1809-1817. doi:10.1016/S0142-9612(01)00307-6
216. Lee TM, Yang CY, Chang E, Tsai RS. Comparison of plasma-sprayed hydroxyapatite coatings and zirconia-reinforced hydroxyapatite composite coatings: In vivo study. *J Biomed Mater Res*. 2004;71A(4):652-660. doi:10.1002/jbm.a.30190
217. Tambasco de Oliveira P, Nanci A. Nanotexturing of titanium-based surfaces upregulates expression of bone sialoprotein and osteopontin by cultured osteogenic cells. *Biomaterials*. 2004;25(3):403-413. doi:10.1016/S0142-9612(03)00539-8

218. Webster TJ, Ejiófor JU. Increased osteoblast adhesion on nanophase metals: Ti, Ti6Al4V, and CoCrMo. *Biomaterials*. 2004;25(19):4731-4739. doi:10.1016/j.biomaterials.2003.12.002
219. Di Iorio D, Traini T, Degidi M, Caputi S, Neugebauer J, Piattelli A. Quantitative evaluation of the fibrin clot extension on different implant surfaces: An *in vitro* study. *J Biomed Mater Res*. 2005;74B(1):636-642. doi:10.1002/jbm.b.30251
220. Papalexioú V, Novaes AB, Grisi MFM, Souza SLS, Taba M, Kajiwará JK. Influence of implant microstructure on the dynamics of bone healing around immediate implants placed into periodontally infected sites. A confocal laser scanning microscopic study. *Clin Oral Implants Res*. 2004;15(1):44-53. doi:10.1111/j.1600-0501.2004.00995.x
221. Iezzi G, Degidi M, Scarano A, Perrotti V, Piattelli A. Bone Response to Submerged, Unloaded Implants Inserted in Poor Bone Sites: A Histological and Histomorphometrical Study of 8 Titanium Implants Retrieved From Man. *Journal of Oral Implantology*. 2005;31(5):225-233. doi:10.1563/1548-1336(2005)31[225:BRTSUI]2.0.CO;2
222. Wisbey A, Gregson PJ, Tuke M. Application of PVD TiN coating to Co-Cr-Mo based surgical implants. *Biomaterials*. 1987;8(6):477-480. doi:10.1016/0142-9612(87)90085-8
223. Scarano A, Piattelli M, Vrespa G, Petrone G, Iezzi G, Piattelli A. Bone Healing around Titanium and Titanium Nitride-Coated Dental Implants with Three Surfaces: An Experimental Study in Rats. *Clin Implant Dent Rel Res*. 2003;5(2):103-111. doi:10.1111/j.1708-8208.2003.tb00191.x
224. Scarano A, Piattelli M, Vrespa G, Caputi S, Piattelli A. Bacterial adhesion on titanium nitride-coated and uncoated implants: an *in vivo* human study. *The Journal of Oral Implantology*. 2003;29(2):80-85. doi:10.1563/1548-1336(2003)029<0080:BAOTNA>2.3.CO;2
225. Groessner-Schreiber B, Neubert A, Müller WD, Hopp M, Griepentrog M, Lange KP. Fibroblast growth on surface-modified dental implants: An *in vitro* study: Fibroblasts on Dental Implant Surfaces. *J Biomed Mater Res*. 2003;64A(4):591-599. doi:10.1002/jbm.a.10417
226. Größner-Schreiber B, Griepentrog M, Hausteín I, et al. Plaque formation on surface modified dental implants: An *in vitro* study. *Clinical Oral Implants Research*. 2001;12(6):543-551. doi:10.1034/j.1600-0501.2001.120601.x
227. Groessner-Schreiber B, Hannig M, Duck A, Griepentrog M, Wenderoth DF. Do different implant surfaces exposed in the oral cavity of humans show different biofilm compositions and activities? *Eur J Oral Sci*. 2004;112(6):516-522. doi:10.1111/j.1600-0722.2004.00171.x
228. Park CY, Kim SG, Kim MD, Eom TG, Yoon JH, Ahn SG. Surface Properties of Endosseous Dental Implants After NdYAG and CO₂ Laser Treatment at Various Energies. *Journal of Oral and Maxillofacial Surgery*. 2005;63(10):1522-1527. doi:10.1016/j.joms.2005.06.015
229. Hallgren C. An *in vivo* study of bone response to implants topographically modified by laser micromachining. *Biomaterials*. 2003;24(5):701-710. doi:10.1016/S0142-9612(02)00266-1
230. Citeau A, Guicheux J, Vinatier C, et al. *In vitro* biological effects of titanium rough surface obtained by calcium phosphate grid blasting. *Biomaterials*. 2005;26(2):157-165. doi:10.1016/j.biomaterials.2004.02.033
231. Schopper C, Ziya-Ghazvini F, Goriwoda W, et al. HA/TCP compounding of a porous CaP biomaterial improves bone formation and scaffold degradation—A long-term histological study. *J Biomed Mater Res*. 2005;74B(1):458-467. doi:10.1002/jbm.b.30199
232. Wen HB, de Wijn JR, Cui FZ, de Groot K. Preparation of calcium phosphate coatings on titanium implant materials by simple chemistry. *J Biomed Mater Res*. 1998;41(2):227-236. doi:10.1002/(SICI)1097-4636(199808)41:2<227::AID-JBM7>3.0.CO;2-K
233. Yang Y, Kim KH, Ong JL. A review on calcium phosphate coatings produced using a sputtering process—an alternative to plasma spraying. *Biomaterials*. 2005;26(3):327-337. doi:10.1016/j.biomaterials.2004.02.029
234. Albrektsson T, Wennerberg A. Oral implant surfaces: Part 2—review focusing on clinical knowledge of different surfaces. *Int J Prosthodont*. 2004;17(5):544-564.